

INTRAFAMILIAL HYBRIDIZATION IN MOSSES? AN ENIGMATIC CASE  
IN THE GENUS *PODPERAEA* (HYPNALES, BRYOPHYTA)

ГИБРИДИЗАЦИЯ МЕЖДУ ПРЕДСТАВИТЕЛЯМИ РАЗНЫХ  
СЕМЕЙСТВ У МХОВ? ЗАГАДОЧНЫЙ СЛУЧАЙ  
В РОДЕ *PODPERAEA* (HYPNALES, BRYOPHYTA)

MICHAEL S. IGNATOV<sup>1</sup> & IRINA A. MILYUTINA<sup>2</sup>

МИХАИЛ С. ИГНАТОВ<sup>1</sup>, ИРИНА А. МИЛЮТИНА<sup>2</sup>

Abstract

A new species from China, *Podperaea bairii*, is described. In addition to morphological differences from the second species of the genus, *P. krylovii* (Amlystegiaceae s.l.), *P. bairii* differs in the nrITS1 sequence, which is very similar to that of the genus *Herzogiella* from the phylogenetically distant family Plagiotheciaceae. At the same time, nrITS2 in *P. bairii* is much more similar to Amblystegiaceae than Plagiotheciaceae. This case is the first report of the putative remote hybridization in mosses.

Резюме

Из Китая описан новый вид, *Podperaea bairii*, который, помимо небольших морфологических отличий от второго вида рода, *P. krylovii* (семейство Amlystegiaceae s.l.), отличается еще последовательностью ITS1, которая соответствует роду *Herzogiella* из филогенетически далекого семейства Plagiotheciaceae. При этом последовательность ITS2 у *P. bairii* значительно более сходна с Amblystegiaceae, нежели с Plagiotheciaceae. Данный случай является первым примером возможной отдаленной гибридизации у мхов.

KEYWORDS: Bryophytes, pleurocarpous mosses, Plagiotheciaceae, Amblystegiaceae, *Podperaea*, new species, China, remote hybridization, nrITS

INTRODUCTION

The genus *Podperaea* was established by Iwatsuki & Glime (1984) for one species, originally described as *Campylium krylovii* Podp. The strongly prorate cells and specific ‘compound’ teeth along leaf margin formed by upper end of lower cell and lower end of next upper cell, were found to be quite odd for *Campylium*. Similar teeth are rare in Hypnales, and are well developed, as far as we know, only in *Myurella* and *Platydictya*.

All other species ever placed in the genus *Campylium* (except *C. krylovii*) never have

‘compound’ teeth. This genus in its traditional circumscription included species that are now treated in many genera: *Campylium* s. str., *Campylophyllum*, *Campyliadelphus*, *Campylidium*, and in addition some species were transferred to *Amblystegium*, *Drepanocladus* and *Pseudocampylium* (cf. Hedenäs, 1997; Ochyra et al., 2003; Ignatov et al., 2007; Vanderpoorten & Hedenäs, 2009). None of these genera have sufficient similarity with *Podperaea krylovii*, and none of them have been found related to it in the molecular analysis of Troitsky et al. (2007), the only study where the position of *Podperaea*

<sup>1</sup> – Main Botanical Garden, Russian Academy of Sciences, Botanicheskaya 4, Moscow 127276 Russia – Россия 127276 Москва, Ботаническая, 4, Главный ботанический сад РАН, e-mail: misha\_ignatov@list.ru

<sup>2</sup> – A.N.Belozersky' Research Institute of Physico-Chemical Biology, Moscow State University, Moscow 119991 Russia – Россия 119991 Москва, МГУ, НИИ Физико-химической биологии им. А.Н. Белозерского, e-mail: iramiljutina@yandex.ru

*krylovii* has been tested by phylogenetic methods.

The placement of the genus remains problematic, which is a common case at the moment for many genera of pleurocarpous mosses. Originally it was referred to Hypnaceae that was a repository for pleurocarps with double costa and complete double alternate peristome and without specific characters that would indicate an affinity to families like Neckeraceae, Sematophyllaceae, Hylocomiaceae, etc. However, the type species of the family Hypnaceae, *Hypnum cupressiforme*, is obviously quite unrelated to *Podperaea* (Troitsky et al., 2007).

*Podperaea krylovii* was described from the Russian Far East and later found in a number of places in South Siberia, and in Hokkaido, Japan (Iwatsuki & Glime, 1984), in Liaoning Province of China (Ignatov et al., 1996) and then also in Inner Mongolia Autonomous District of China (Zhao et al., 2006; Bai, 2010).

Collections from different parts of Russia studied by us have been fairly homogeneous morphologically, so posed no questions about their species identity. The variation seen in Chinese specimens however was bigger: specimens from Khingan Mts. (ranging from Amurskaya Province of Russia and Heilongkiang Province of China to Inner Mongolia) were found nearly identical to Russian ones, whereas plants from another region, Inner Mongolia, appeared to be different in morphology, so the testing with molecular markers was applied to these specimens.

An unexpected preliminary results have forced us to search additional collections from Inner Mongolia, and new gatherings have confirmed the preliminary observation that the most widespread population in the Inner Mongolia is an undescribed species, *Podperaea bairii* (see description below).

#### MATERIAL AND METHODS

The taxa selection for phylogenetic analysis was done with the focus on genera possibly related to *Podperaea* according to the results of Troitsky et al. (2007). The subset of taxa used in the previous analyses (Ignatov et al., 2007; Troitsky et al., 2007) was taken. New and already published sequence data used in analysis are given in Appendix 1.

Laboratory protocol was essentially the same as in some of our previous analyses (e.g., Gardiner et al., 2005). The sequences were aligned manually in Bioedit (Hall, 1999). Maximum parsimony analysis and jackknifing for 1000 replications was performed in Nona (Goloboff, 1994) under Winclada shell (Nixon, 1999).

In the present paper only ITS data are analyzed, as this DNA region is the most variable in mosses, and the results of its sequence analyses often correspond to genera and families of many pleurocarpous moss groups in their traditional circumscriptions. Preliminary tests with chloroplast markers *trnL-F* and *rps4* have given poor resolution and are not presented here.

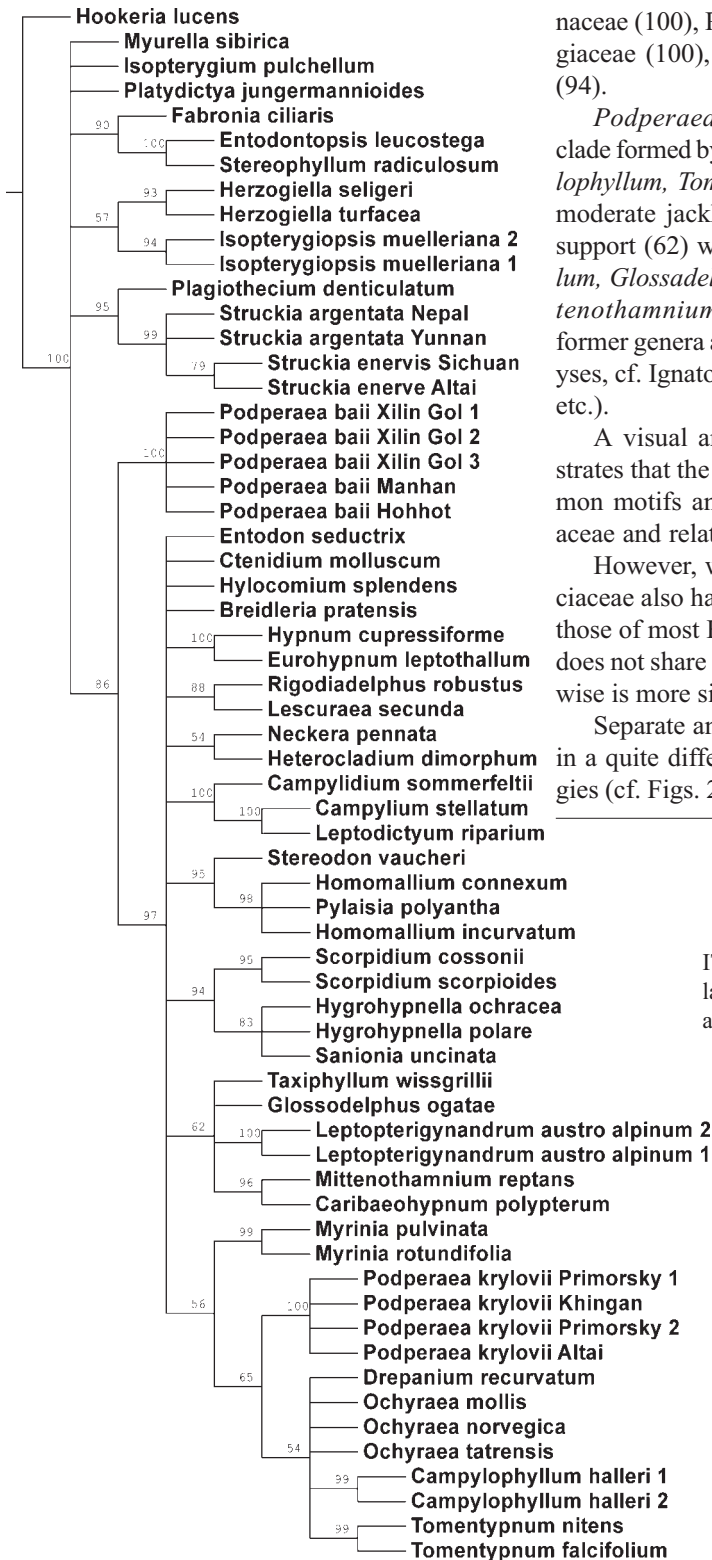
As the preliminary study revealed a considerable difference between systematic position that follows from a separate analysis of ITS1 and ITS2, these loci were studied separately. Hereafter within this paper we will call them ITS1 and ITS2 for simplicity, although the former locus includes ITS1, 5.8 RNA gene and a short part of 3'-end of ITS2 (38 positions in the alignment (30-37 nucleotides for different species, up to 678 position of the alignment), while ITS2 in a slightly reduced part of this spacer, representing 503 positions closer to 5'-end, i.e. without 38 mentioned positions of the alignment from the 3'-end.

#### RESULTS

The MP tree based on the ITS1-2 region and rooted on *Hookeria* (Fig. 1) includes basal polytomy of the species classified in Plagiotheciaceae (sensu Pedersen & Hedenäs, 2002; Ignatov et al., 2007, etc.) and also Fabroniaceae and Stereophyllaceae and terminal clade.

Within the basal polytomy, a high jackknife support was obtained for clades formed by species of one genus, e.g., *Herzogiella* (93), *Struckia* (99), and also *Plagiothecium*+*Struckia* clade was supported (95), as well as Fabroniaceae+*Stereophyllaceae* (90).

Terminal clade includes two subclades. The first small subclade is formed by 5 accessions of *Podperaea bairii* (jackknife support 100). The second larger subclade (support 97) represents a poorly resolved terminal polytomy of various groups of Hypnales. Within this polytomy, a high support has been received by many families (usually represented by few specimens), e.g., Hyp-



naceae (100), Pseudoleskeaceae (88), Amblystegiaceae (100), Pylaisiaceae (95), Scorpidiaceae (94).

*Podperaea krylovii* was found within the clade formed by *Ochyraea*, ‘*Drepanium*’, *Campylophyllum*, *Tomentypnum*. This clade got only a moderate jackknife support, 65. A comparable support (62) was found for a clade of *Taxiphyllum*, *Glossadelphus*, *Leptopterigynandrum*, *Mittenothamnium*, *Caribaeohypnum* (the three former genera appear in many phylogenetic analyses, cf. Ignatov et al., 2007; Olsson et al., 2009, etc.).

A visual analysis of the alignment demonstrates that the ITS1 of *Podperaea bairii* has common motifs and substitutions with Plagiotheciaceae and related families (Fig. 4).

However, within the ITS2, where Plagiotheciaceae also have sequences quite different from those of most Hypnales (Fig. 5), *Podperaea bairii* does not share ‘plagiothecioid’ motifs, but otherwise is more similar to some Amblystegiaceae.

Separate analyses of ITS1 and ITS2 resulted in a quite different strict consensus tree topologies (cf. Figs. 2 and 3). The former one made for

Fig. 1. Jackknife tree based on ITS 1–2 dataset; supports >50% calculated for 1000 replications are shown above branches.

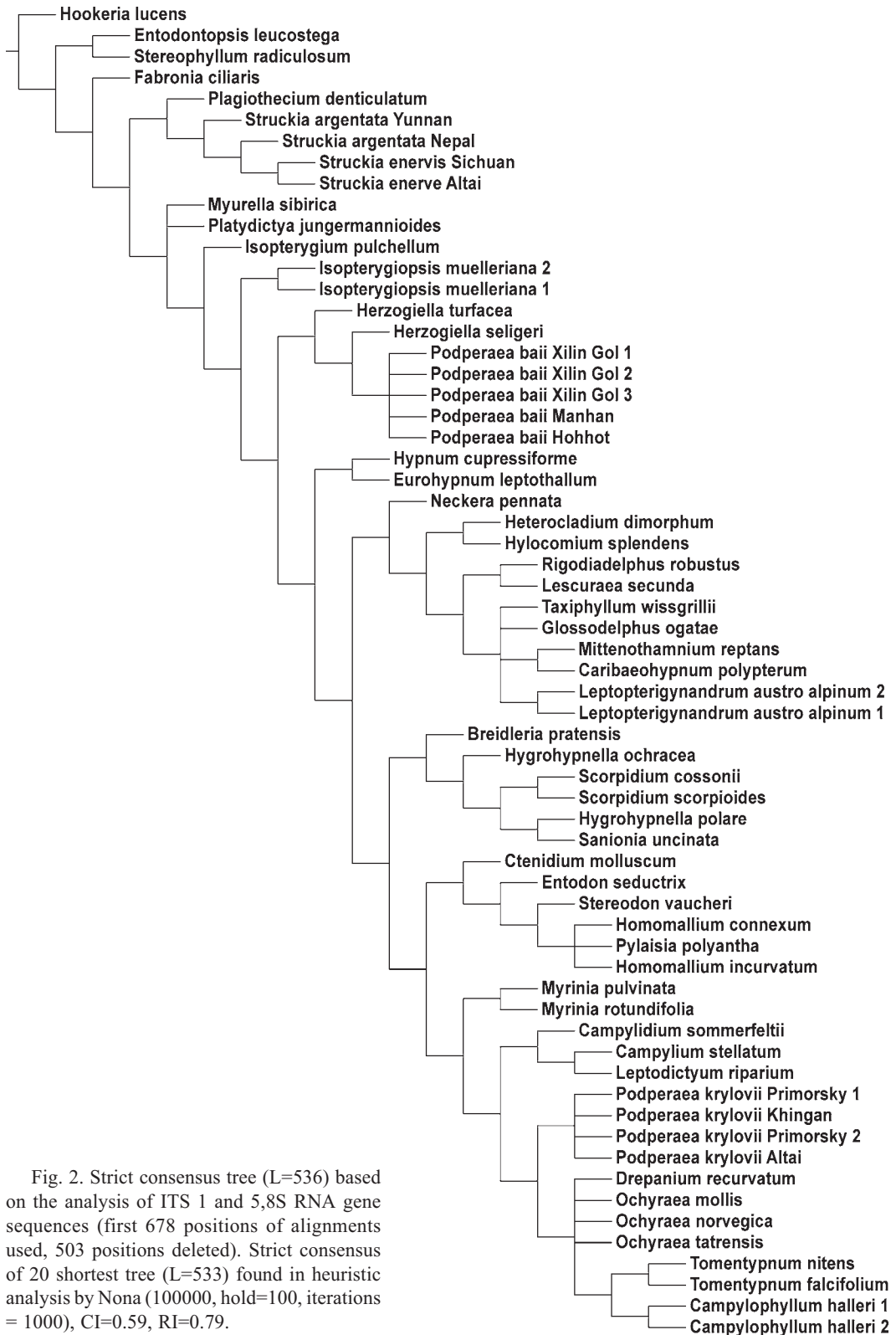


Fig. 2. Strict consensus tree (L=536) based on the analysis of ITS 1 and 5,8S RNA gene sequences (first 678 positions of alignments used, 503 positions deleted). Strict consensus of 20 shortest tree (L=533) found in heuristic analysis by Nona (100000, hold=100, iterations = 1000), CI=0.59, RI=0.79.

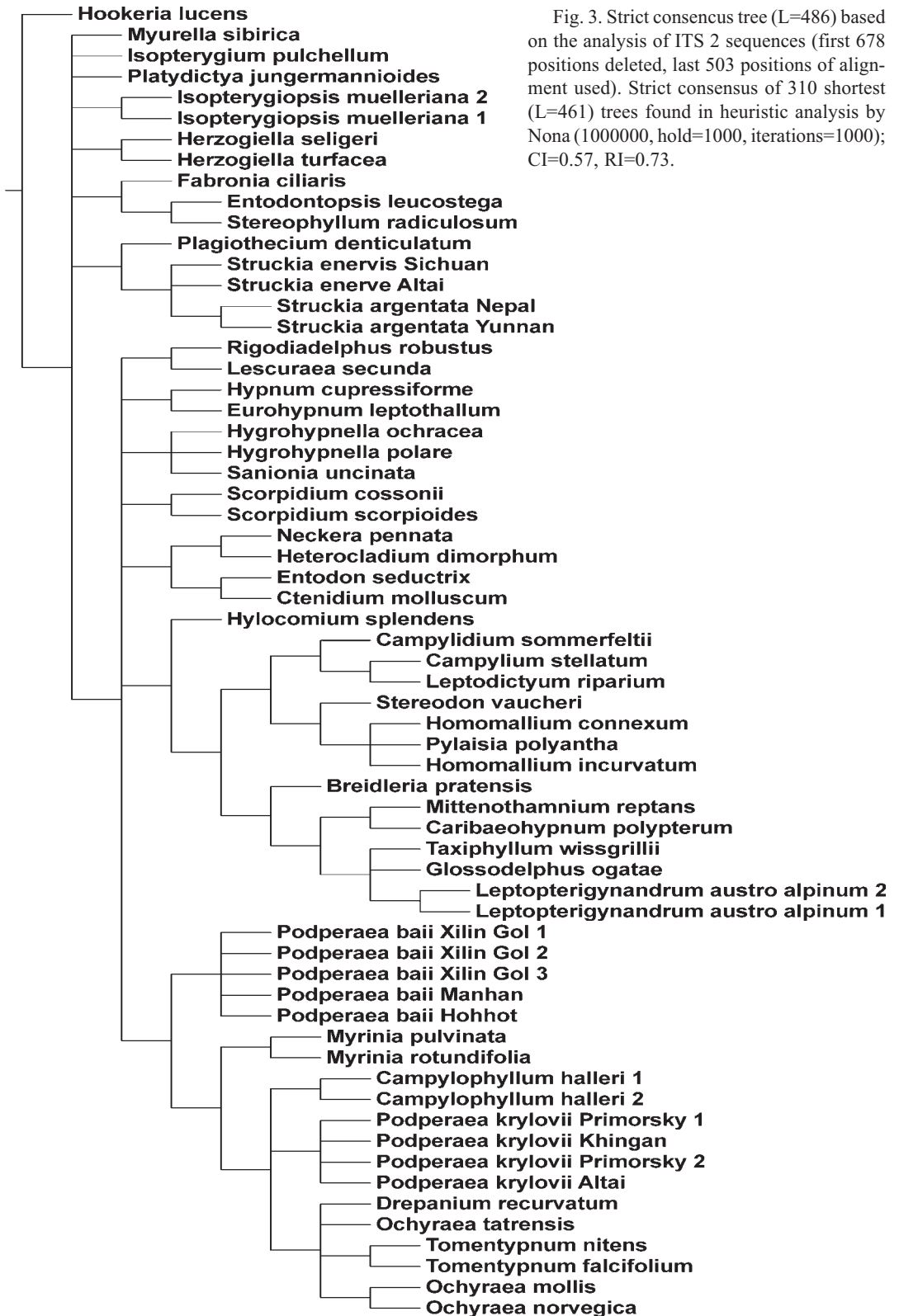


Fig. 3. Strict consensus tree (L=486) based on the analysis of ITS 2 sequences (first 678 positions deleted, last 503 positions of alignment used). Strict consensus of 310 shortest (L=461) trees found in heuristic analysis by Nona (1000000, hold=1000, iterations=1000); CI=0.57, RI=0.73.

	100	110	120	130	140	150	160	170	180	.....
HOMOMALLIUM CONNEXUM	CCTTCG	CCAATCGCCACGCG	CG					CGCGGCGT	CGGG	TT
HOMOMALLIUM INCURVATUM	CCTTCG	CCAATCGCCACGCG	CG					CGCGGCGT	CGGG	TT
PYLAISIA POLYANTHA	CCTTCG	CCAATCGCCACGCG	CG					CGCGGCGT	CGGG	TT
STEREBODON VAUCHERI	CCTTCG	C	CACGCG	CG				CGCGGCGT	CGGG	TT
HYPNUM CUPRESSIFORME	CCTTCG	T	CACGAG	C	TC	GCT		CGTGGCGC	GGGG	CT
EUROHYPNUM LEPTOTHALLUM	CCTTCG	T	CACGAG	C	TC	GCT		CGTGGCGC	GGGG	CTT
BREIDLERIA PRATENSIS	CCTTCG	T	CACGA	CGAGC		GC	GCTCGCT	CGGAGGT	CGGG	TT
CTENIDIUM MOLLUSCUM	CCTTCG	T	CACGA	CGAGC	T			CGT	CGGT	GGGG
NECKERA PENNATA	CCT	CG	T	CACGCG	C	TC	GC	CGTGGCGT	CGGG	TT
HYLOCOMIUM SPLENDENS	CCTTCG	T	CACGT	C	TC		GGGA	CGTGGCGA	AGGGGG	AT
ENTODON SEDUCTRIX	CCTTCG	T	CACGAG	CGAGC	TC			CGCGGCGT	GGGG	TT
MITTENOTHAMNIUM REPTANS	CCTTCG	T	CACGAG	C			AAG ACT	CGGGGCGT	GGGG	TT
HYGROHYPNELLA OCHRACEA	CCTTCG	T	CACGGG	CGAGA			GCT	CGGGGCGT	CGGG	TT
HYGROHYPNELLA POLARE	CCTTCG	T	CACGAG	CGAGA			GCT	CGCGGCGT	CGGG	TT
SANIONIA UNCINATA	CCTTCG	T	CACGAG	GTCTGA			CCT	CGCGGCGT	CGGG	TT
SCORPIDIUM COSSONII	CCTTCG	T	CACGAG	CGAGA			GCT	CGGGGCGT	CGGG	TT
SCORPIDIUM SCORPIOIDES	CCTTCG	T	CACGAG	CGAGA			GCT	CGGGGCGT	CGGG	TT
CARIBABOHYPNUM POLYPTERUM	CCTTCG	T	CGGAG	C	TCGG	TCGA	GTC	ACGGA	CGGGGCGT	CGGG
HETEROCLADUM DIMORPHUM	CCTTCG	T	CACGAG	C	TTGG	TC		ACGA	CGGGGCGT	CGGG
LESCURAEA SECUNDA	CCTTCG	T	CACGAG	C	T			CGTGGCGT	CGGG	TT
RIGODIADELPHUS ROBUSTUS	CCTTCG	T	CACGAG	C	TTGG	TC		ACGA	CGGGGCGT	CGGG
TAXIPHYLUM WISSGRILLII	CCTTCG	T	CACGAG	C	TTGG	TC	ACGTC	ACGA	CGGGGCGT	CGGG
LEPTOPTERIGYNANDRUM A-ALPINUM2	CCTTCG	T	CACGAG	C	TTGG	TCT	CGTC	ACGA	CGGAGGT	CGGG
LEPTOPTERIGYNANDRUM A-ALPINUM1	CCTTCG	T	CGGAG	C	TTGG	TCC	CGTC	ACGA	CGGGGCGT	CGGG
GLOSSODELPHUS OGATAE	CCTTCG	T	CACGAG	C	TTGG	TC	ACGTC	ACGA	CGGGGCGT	CGGG
DREPANIUM RECURVATUM	CCTTCG	T	CACGAG	CGAGC	TC	CCCTC	GGCA	GCTCCCT	CGCGGCGT	CGGG
OCHYRAEA MOLLIS	CCTTCG	T	CACGAG	CGAGC	TC	CCCTC	GGCA	GCTCCCT	CGCGGCGT	CGGG
OCHYRAEA NORVEGICA	CCTTCG	T	CACGAG	CGAGC	TC	CCCTC	GGCA	GCTCCCT	CGCGGCGT	CGGG
OCHYRAEA TATRENSIS	CCTTCG	T	CACGAG	CGAGC	TC	CCCTC	GGCA	GCTCCCT	CGCGGCGT	CGGG
CAMPYLOPHYLLUM HALLERI_1	CCTTCG	T	CACGAG	CGAGC	TC		GGCA	GCTCTCT	CGCGGCGT	CGGG
CAMPYLOPHYLLUM HALLERI_2	CCTTCG	T	CACGAG	CGAGC	TC		GGCA	GCTCTCT	CGCGGCGT	CGGG
MYRINIA PULVINATA	CCTTCG	T	CACGCT	CGAGC	TC			CGTGGCGT	CGGG	TT
MYRINIA ROTUNDIFOLIA	CCTTCG	T	CACGCT	CGAGC	TC			CGTGGCGT	CGGG	TT
CAMPYLUM STELLATUM	CCTTCG	T	CACGCG	CGAGCGAGTTC	CTC	GTGG		GCTCCCTCGCGGCGCGGCGT	CGGG	T
LEPTODICTYUM RIPARIUM	CCTTCA	T	CACG	G	AGAGCGAGTTC	CTC	TGG	GCTCCCTCGCGGCGCGGCGT	CG	G
CAMPYLIDIUM SOMMERFELTII	CCTTCG	T	CACGCG	CGAGCGAG	TC	CCCTC	GTGG	GCTCCCTCGCGGCGCGGCGT	CGGG	T
TOMENTYPNUM NITENS	CCTTCG	T	CACGAG	CGAGCGAGCTC	TC	GGCA		GCTCTCT	CGCGGCGT	CGGG
TOMENTYPNUM FALCIFOLIUM	CCTTCG	T	CACGAG	CGAGCGAGCTC	TC	GGCA		GCTCTCT	CGCGGCGT	CGG
PODPERAEA KRYLOVII PRIMORSKY1	CCTTCGGT	T	CGCGCG	CGAGCCCGCTC		GGCG		GCTCCCT	CGCGGCGT	CGGG
PODPERAEA KRYLOVII KHINGAN	CCTTCG	T	CACGCG	CGAGCCCGCTC		GGCG		GCTCCCT	CGCGGCGT	CGGG
PODPERAEA KRYLOVII PRIMORSKY2	CCTTCG	T	CACGCG	CGAGCCCGCTC		GGCG		GCTCCCT	CGCGGCGT	CGGG
PODPERAEA KRYLOVII ALTAI	CCTTCG	T	CACCGC	CGAGCCCGCTC		GGCG		GCTCCCT	CGCGGCGT	CGGG
PODPERAEA BAII_XILIN_GOL1	TCGAGG	T	CACGAG	C	GT			CAAAACT	CGCGGCGT	CGGGGG
PODPERAEA BAII_XILIN_GOL2	TCGAGG	T	CACGAG	C	GT			CAAAACT	CGCGGCGT	CGGGGG
PODPERAEA BAII_XILIN_GOL3	TCGAGG	T	CACGAG	C	GT			CAAAACT	CGCGGCGT	CGGGGG
PODPERAEA BAII_MANHAN	TCGAGG	T	CACGAG	C	GT			CAAAACT	CGCGGCGT	CGGGGG
PODPERAEA BAII_HOHOT	TCGAGG	T	CACGAG	C	GT			CAAAACT	CGCGGCGT	CGGGGG
PLAGIOTHECIUM DENTICULATUM	CCGAAG	T	CACGAG	C	GT			CAAAACT	CGCGGCGT	CGGGGG
STRUCKIA ENERVIS_SICHUAN	CCGATG	T	CTCGAG	C	GT			CAAAACT	CGGGGCGT	CGGGGG
STRUCKIA ARGENTATA_NEPAL	CCGAAG	T	CTCGAG	C	GT			CAAAACT	CGGGGCGT	CGGGGG
MYURELLA SIBIRICA	CCGAGG	T	CACGAG	C	GT			CAAAACT	CGTGGCGT	CGGGGG
HERZOGIELLA SELIGERI	TCGAGG	C	CCGAGC	C	GT			CAAAACT	CGGGGCGT	CGGGGG
HERZOGIELLA TURFACEA	TCGAGG	T	CACGAG	C	GT			CAAAACT	CGGGGCGT	CGGGGG
STRUCKIA ARGENTATA_YUNNAN	CCGAGT	T	CTCGAG	C	GT			CAAAACT	CGCGGCGT	CGGGGG
STRUCKIA ENERVE_ALTAI	CCGATG	T	CTCGAG	C	GT			CAAAACT	CGGGGCGT	CGGGGG
ISOPTERYGIOPSIS MUELLERIANA2	CCGAGG	T	CACGAG	C	GT			CGCAACT	CGTGGCGT	CGGGGG
ISOPTERYGIOPSIS MUELLERIANA1	CCGAGG	T	CACGAG	C	GT			CGCAACT	CGTGGCGT	CGGGGG
ISOPTERYGIUM PULCHELLUM	CCGTTG	T	CACGAG	C	GT			CGCAACT	CGTGGCGT	CGGGGG
PLATYDICTYA JUNGERMANNIOIDES	CCGAGT	T	CACGAG	C	TT			CAAAACT	CGTGGCGT	CGGGGG
ENTODONTOPSIS LEUCOSTRGA	CCGAGG	T	CGCGAG	C			CCGA	CT	CGCGGCGT	CGGGGG
STERBOPHYLLUM RADICULOSUM	CCGAGG	T	CGCGAG	C			CCGA	CT	CGCGGCGT	CGGGGG
FABRONIA CILIARIS	CCGAGA	C	CGCGAG	C	TT			CAAACT	CACGGCGT	CGGGGG
HOOKERIA LUCENS	CCGAGG	T	CACGAGA	C	TT			CCC	GCT	CGCGGTTCCGGGGGA

Fig. 4. Part of ITS 1 alignment, showing common substitutions and indels of *Podperaea baii* and Plagiotheciaceae.

ITS1, resolved *Podperaea baii* as sister to *Herzogiella* within the basal grade.

At the same time, strict consensus for ITS2 sequences has found both *Podperaea krylovii* and *P. baii* in one clade, which also includes *Myriniia* and four genera, *Ochyraea*, ‘*Drepanium*’, *Campylophyllum*, *Tomentypnum*, the same as have been found in the combined analysis of ITS1 and ITS2.

Visual evaluation of sequences demonstrates at once that ITS1 region in *Podperaea baii* has all characteristic motifs of Plagiotheciaceae, which are numerous and readily seen (Fig. 4).

ITS2 in pleurocarpous mosses is less variable compared with ITS1 and rather few characteristic places in the alignment are revealing. Most of them demonstrate the difference between

	690	700	710	1080	1090	1100	1110	1120	1130	1140	
HOMOMALLIUM_CONNEXUM	TC		GGTC	CGCGCCC	TT	CATG				GC	GC
PYLAIASIA_POLYANTHA	TC		GGGG	CGT	GCCCC	TT	CAT			GC	GGC
HOMOMALLIUM_INCURVATUM	TC		GGGG	CGCGCCCC	TT	CATG				GC	GGC
STEREODON_VAUCHERI	GTG		AACCGGGG	CGCGCCCC						GC	GGC
TAXIPHYLUM_WISSGRILLII	AGC		CGC	CGCGCCCC						GC	GGC
LEPTOPTERIGYNANDRUM_A-ALPINUM2	T		CGC	CGCGCCCC						GC	GGC
LEPTOPTERIGYNANDRUM_A-ALPINUM1	T		CGC	CGCGCCCC						GC	GGC
GLOSSODELPHUS_OGATAE	AGT		CGC	CGCGCCCC						GC	GGC
ENTODON_SEDUCTRIX	GCC		GGG	TCACGCCCCCCC						GT	GGC
MITTENOTHAMNIUM_REPTANS				CGCGCCCC						GC	GGC
CARIBAOHYPNUM_POLYPTERUM	GCCCCGCGCG		CGC	CGCGCCCC						CGCGGGC	GGC
HYPNUM_CUPRESSIFORME	GTT	CGCG	GGG	CGCGCCC						GCGA	GGC
EUROHYPNUM_LEPTOTHALLUM	GTT	CGCG	GTTG	CGCGCCCC						GCGA	GGC
CTENIDIUM_MOLLUSCUM	GTG		GGG	CGCGCCCC						GCGG	GTT
NECKERA_PENNATA	ACC		GGGA	CGCGCCCC						GC	GGC
HETEROCLADIUM_DIMORPHUM	ACC		GGG	CGCGCCCC						GC	GGC
HYLOCOMIUM_SPLENDENS			GGG	CG	GCCCCC					GGC	GGC
BREIDLERIA_PRATENSIS	GCC		GGG	CGCGCCCC						GC	GGC
CAMPYLOPHYLLUM_HALLERI_1	ATT		CATTGGG	CGCGCCCC						GC	GGC
CAMPYLOPHYLLUM_HALLERI_2	ATT		CATTGGG	CGCGCCCC						GC	GGC
DREPANIUM_RECURVATUM	TTT		GGG	CGCGCCCC						GC	GGC
CAMPYLUM_STELLATUM	GTG		AAATCGGG	CGCGCCACCCCC	CCCG	CATGGGTTTGG				GC	GGC
LEPTODICTYUM_RIPARIUM	GTGT		GAATCGGG	CGC	CCACCCCC	CCCG	CATGGGTTTGG			GC	GGC
HYGROHYPNELLA_OCHRACEA	GTC		GGG	CGCGCCCC						GT	GGC
HYGROHYPNELLA_POLARE	GCC		GGG	CGCGCCCC						GC	GGC
RIGODIADELPHUS_ROBUSTUS	GCG		CGC	CGCGCCAC						GC	GGC
SANIONIA_UNCINATA	GCC		GGG	CGCGCCCC						GC	GGC
SCORPIDIUM_COSSONII	GCC		GGG	CGCGTCCCC						GT	GGC
SCORPIDIUM_SCORPIOIDES	GCC		AGG	CGCGTCCCC						GT	GGC
CAMPYLIDIUM_SOMMERFELTII	GCC		GGG	CGCGCC						GC	GGC
LESCURAEA_SECUNDA	GTG		TCG	CGCGCCCC						GT	GGC
OCHYRAEA_MOLLIS	CTT		GGG	CGCGCCCC						GT	GGC
OCHYRAEA_NORVEGICA	CTT		GGG	CGCGCCCC						GT	GGC
OCHYRAEA_TATRENSIS	TT		GGG								
TOMENTYPNUM_NITENS	CTT		CGGG	CGCGCCCC						GC	GGC
TOMENTYPNUM_FALCIPOLIUM	CTT		CGGG	CGCGCCCC						GC	GGC
MYRINIA_PULVINATA	CTT		G	CGCGCCCC						GC	GGC
MYRINIA_ROTUNDFOLIA	CTT		GATCGGG	CGCGCCCC						GC	GGC
PODPERAEA_KRYLOVII_PRIMORSKY_1	CTT		GATCGGG	CGCGCCCC						GC	GGC
PODPERAEA_KRYLOVII_KHINGAN	CTT		GATCGGG	CGCGCCCC						GC	GGC
PODPERAEA_KRYLOVII_PRIMORSKY_2	CTT		GATCGGG	CGCGCCCC						GC	GGC
PODPERAEA_KRYLOVII_ALTAI	CTT		GATCGGG	CGCGCCCC						GC	GGC
PODPERAEA_BAII_HELAN_1	CTT		GTGAATCGGG	CGCGCCCC						GC	GGC
PODPERAEA_BAII_HELAN_2	CTT		GTGAATCGGG	CGCGCCCC						GC	GGC
PODPERAEA_BAII_HELAN_3	CTT		GTGAATCGGG	CGCGCCCC						GC	GGC
PODPERAEA_BAII_MANHAN	CTT		GTGAATCGGG	CGCGCCCC						GC	GGC
PODPERAEA_BAII_HOHHOT	CTT		GTGAATCGGG	CGCGCCCC						GC	GGC
PLAGIOTHECIUM_DENTICULATUM	A		GCG	C	GTCCCC					TTGGCGCTCC	TT
STRUCKIA_ENRIVIS_SICHUAN	G		GCG	C	GTCCCC					TTGGCGCTCC	TT
STRUCKIA_ARGENTATA_NEPAL	G		GCG	C	GTCCCC					TTGGCGCTCC	TT
MYURELLA_SIBIRICA	A		GCG	C	CGC					TTGGCGCTCT	TC
HERZOGIELLA_SELIGERI	A		GCG	C	GCCCC					TTGGCGCTCC	TT
HERZOGIELLA_TURFACEA	A		GCG	C	GCCCC					TTGGCGCTCC	TT
STRUCKIA_ARGENTATA_YUNNAN	G		GCG	C	GTCCCC					TTGGCGCTCC	TT
STRUCKIA_ENERVE_ALTAI	G		GCG	C	GTCCCC					TTGGCGCTCC	TT
ISOPTERYGIOPSIS_MUELLERIANA_2	G		GCG	C	GCCCC					TTGGCGCTCC	TT
ISOPTERYGIOPSIS_MUELLERIANA_1	C		GCG	CGCGCCCC						TTGGCGCTCC	TT
ISOPTERYGIUM_PULCHELLUM	A		GCG	C	GCCCC					TTGGCGCTCC	TT
PLATYDICTYA_JUNGERMANNIOIDES	A		GCG	C	CCCC					TTGGCGCTCC	TT
ENTODONTOPSIS_LEUCOSTEGA	G		GCG	C	CTCC					TCGGCGCTCC	TT
STEREOPHYLLUM_RADICULOSUM	A		GCG	C	CTCC					TCGGCGCTCC	TT
FABRONIA_CILIARIS	A		GCG	C	CCCC					TCGGCGCTCC	TT
HOOKERIA_LUCENS	A		GCG	C	CCCC					TCGGCGCTCC	TT

Fig. 5. Part of ITS 2 alignment, showing absence of common substitutions and indels of *Podperaea baii* and Plagiotheciaceae.

Plagiotheciaceae+Stereophyllaceae+Fabroniaceae+Hookeriaceae and the rest of group represented in the analysis. However none of them indicate the relationship of *P. baii* with Plagiotheciaceae, but instead show that it belongs to that subclade of the terminal polytomy in Fig. 1 where *Podperaea krylovii*, *Campylophyllum*, *Ochryaea*, *Myrinia* and *Tomentypnum* occur.

When the first sequence of *P. baii* was analyzed, the possible contamination was suspected. Thus the DNA was cloned and re-sequenced and also re-extracted from sporophyte tissue of the same collection. An additional collections from Inner Mongolia were also looked for, including a special field trip in 2011, and two of new collections of *P. baii*, from geographically different

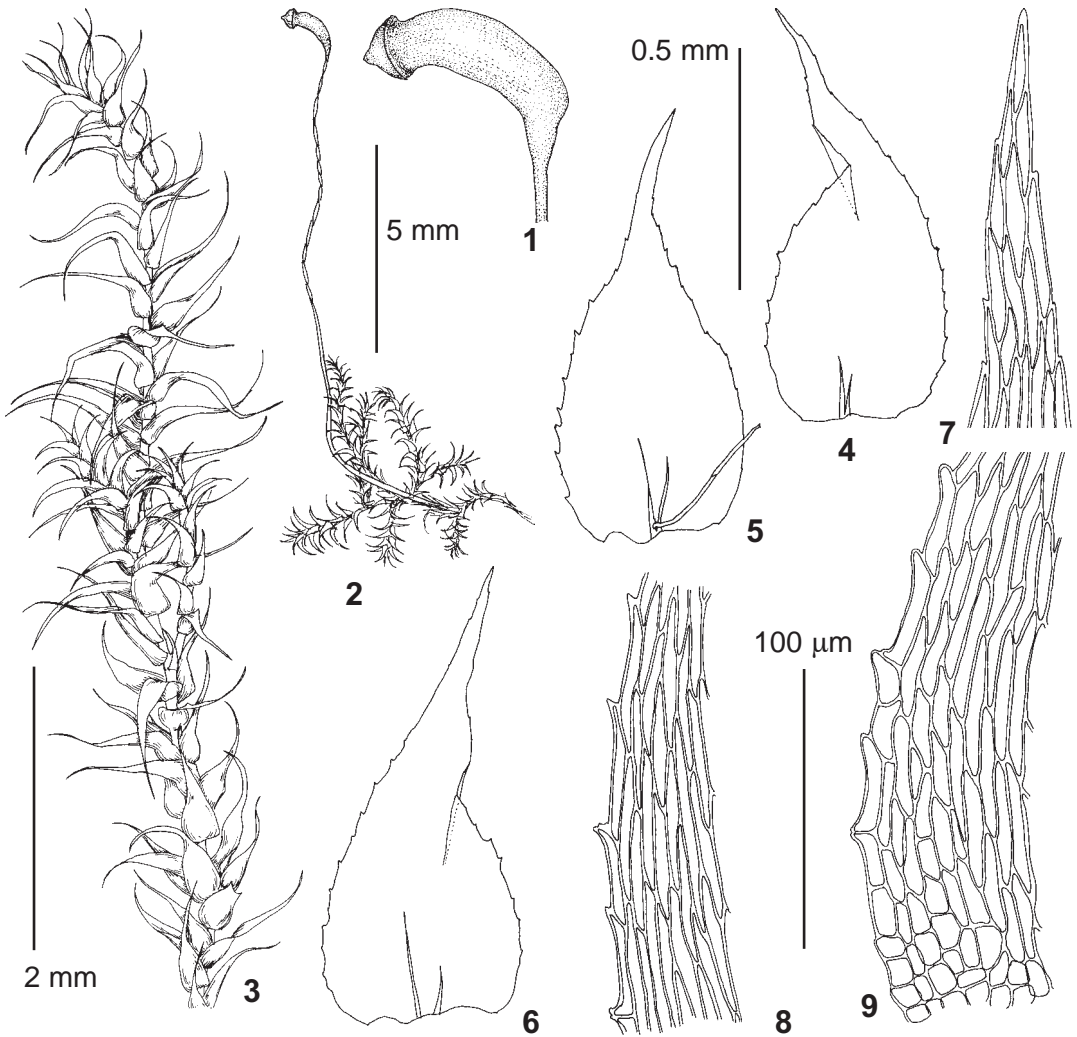


Fig. 6. *Podperaea baii* (Hedw.) Müll. Hal. (from isotype: China, Inner Mongolia, Xilin Gol, Bai X.-L. 058141, MHA): 1 – capsule; 2–3 – habit; 4–6 – leaves; 7 – upper laminal cells; 8 – mid-leaf cells; 9 – basal laminal cells. Scale bars: 5 mm for 2; 2 mm for 1, 3; 0.5 mm for 4–6; 100 µm for 7–9.

areas within Inner Mongolia Autonomous District of China have given the same ITS sequence: “plagiothecoid” ITS1 and “non-plagiothecoid” ITS2.

#### DISCUSSION

The ITS and especially ITS1 of Plagiotheciaceae–Stereophyllaceae–Fabroniaceae group has many specific motifs (Fig. 4), also shared with Hookeriaceae. The mentioned families are characterized by expanded insertions, as well as substitutions in a fairly conservative places (Fig. 4).

Peculiar motifs of Plagiotheciaceae in ITS2

are not so many, but nevertheless some expanded indels differ this family (and some above mentioned related families) from the rest of Hypnales (Fig. 5). However none of them are found in *P. baii*, in contrast with ITS1 were the ‘plagiothecoid specificity’ is totally shared by *P. baii*.

ITS2 of *P. baii* differs somewhat also from *P. krylovii*, although strict consensus tree in the maximum parsimony analysis resolves them within one clade, together with marginal groups of Amblystegiaceae (*Camylophyllum*, *Tomentypnum*, *Drepanium*, and *Myrinia*).



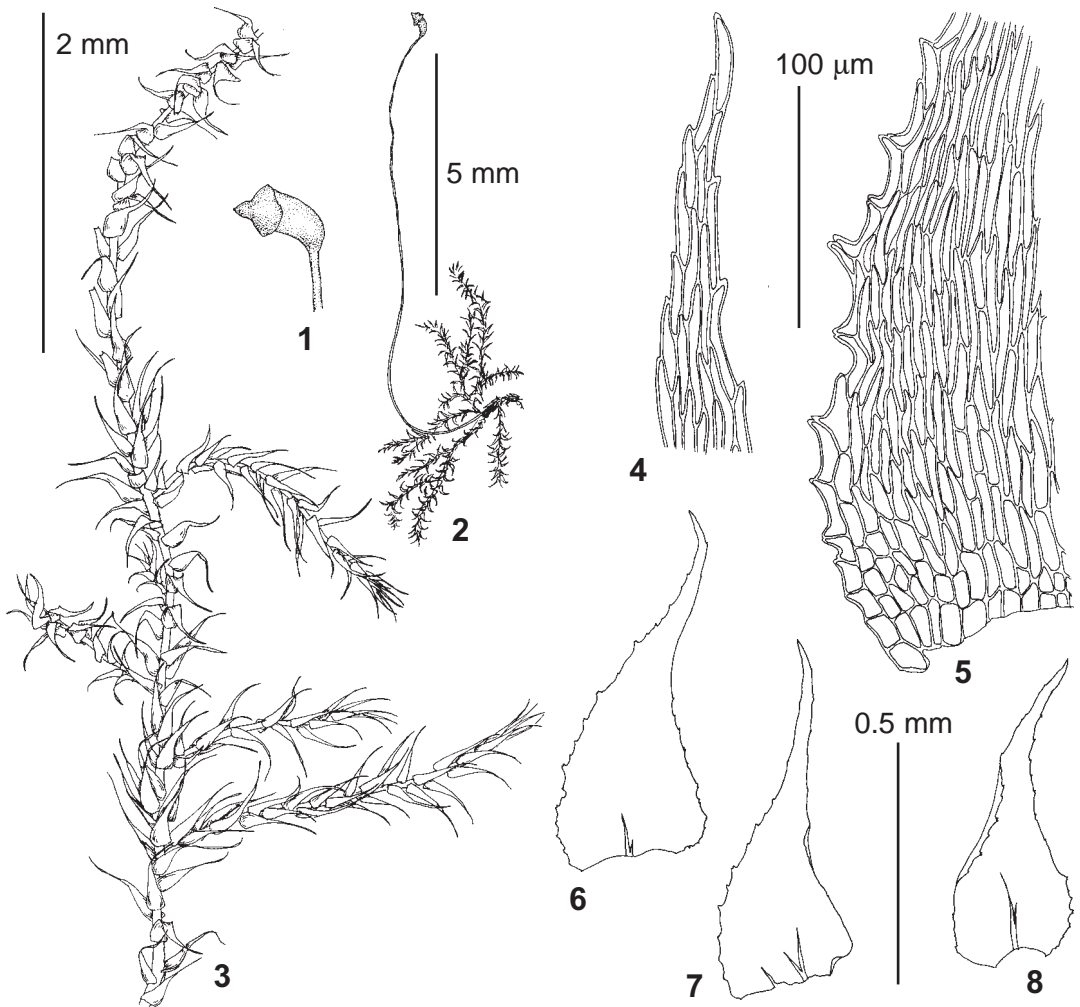


Fig. 7. *Podperaea krylovii* (Podp.) Müll. Hal. (from China, Inner Mongolia, Khingan Mts., Bai X.-L. 04095, MHA): 1 – capsule; 2-3 – habit; 4 – upper laminal cells; 5 – mid-leaf and basal laminal cells; 6-8 – leaves. Scale bars: 5 mm for 2; 2 mm for 1, 3; 0.5 mm for 6-8; 100 µm for 4-5.

Morphological differences between Plagiotheciaceae and ‘Hypnales without Plagiotheciaceae and related families’ include a number of characters and many of them are highly diagnostic. However, the genus *Herzogiella* does not belong to the ‘core Plagiotheciaceae’, so the most unique characters of the family, e.g., axillary position of rhizoides, ‘lack of pseudopraphyllia’ (cf. Ignatov & Hedenäs, 2007), axillary 3-4-celled gemmae, and lack of regular pinnate branching are not the case of this genus. Other characters, like pale peristome and a tendency to a complanate branching being common in Plagiotheciaceae, are known in a many other pleurocarpous groups. Molecular phylogenetic studies however

always resolved *Herzogiella* in Plagiotheciaceae, excepting a likely misplaced in the genus *H. adscendens* (Lindb.) Z. Iwats. & W.B. Schofield.

Although the morphological circumscription of *Herzogiella* is rather indefinite, this genus is distinct from *Podperaea* in having only simple marginal teeth, while teeth in the latter are ‘compound’. The presence of such teeth in *Podperaea bairii* is the main reason for its placement in this genus.

#### ***Podperaea bairii* Ignatov sp. nov.**

*A Podperaea krylovii* plantis robustioribus, foliis longioribus et latioribus, cellulis latoiribus, glabris, sporiis magnis, et ITS sequentia valde differt.

**Holotype:** China, Inner Mongolia (NE, desert area), Xilin Gol, Hun shan de ka, 21.VIII.2005, coll. Bai X.-L. # 058141 (holotype MHA, isotype HIMC).

**Ethymology:** the species is named in honor of Prof. Bai Xue-liang, an eminent bryologist, who published moss flora of Inner Mongolia in China and collected this moss species.

Plants small, in loose, light green mats. Stem prostrate, to 1 cm long, with weak central strand, without hyalodermis, irregularly to rather regularly pinnately branched. Proximal branch leaves lanceolate to ovate, sitting around branch primordia on stem. Leaves ovate-lanceolate, gradually more or less longly acuminate, 0.7-0.95×0.35-0.45 mm, serrate all around by simple and partly 'compound' teeth; 'compound' teeth rather perpendicular to leaf margin, and more or less acute; costa double or forked, short and weak, or sometimes one of branches of forked costa extends to 0.5 the leaf length; laminal cells 30-70×6-7 µm; basal cells quadrate to short rectangular, 7-11 µm wide, forming small indistinctly delimited group. Autoicous. Perichaetial leaves somewhat larger than stem leaves, with single costa reaching up half-leaf, serrulate, smooth. Setae to 15 mm. Capsules ca. 1.3 mm long, arcuate and strongly contracted below the mouth when dry; operculum conic; annulus separated by fragments; peristome double, complete, with long cilia. Spores 17-18 µm.

**Specimens examined:** CHINA, INNER MONGOLIA: Xilin Gol, Bai X.-L. #058141 (HIMC, MHA); Manhan Shan, Ignatov #11-1 (MHA); Hohhot, Kou Jin #110(7) (HIMC, MHA).

We did not see specimens from Helan Shan, alt. 1900 m, HQ269, but according to illustrations in Bai (2010) and Zhao et al. (2006) it is.

**Distinction.** *Podperaea bairii* is a somewhat larger plant than *P. krylovii*, it has broader laminal cells without prorate upper angles; 'compound teeth' are 'more acute' as cells forming a tooth join each other up to the tooth apex, whereas in *P. krylovii* cells forming a tooth are more or less divergent distally, at least at short distance (cf. Figs. 6 & 7). In addition, spores in *P. bairii* are larger than in *P. krylovii*, 17-18 µm versus 10-12 µm.

\* \* \*

Remote hybridization is well-known in flowering plants, where widespread artificial and occasionally natural intrageneric hybrids are known (*Triticum*×*Secale* = *Triticale*; *Hordem*×*Elymus* = *Hordelymus*; *Sorbus*×*Cotoneaster* = *Sorbocotoneaster*, etc.). These and similar cases however include different genera that belong to the same subfamily, and thus represent a hybridization of only a moderately divergent groups.

Much more remote hybridization is known in orchids, where two or more not very closely related genera may produce stable hybrids: *Cattleya* × *Laelia* = *Laeliocattleya*; *Cattleya* × *Sophranitis* = *Sophranocattleya*; *Cattleya* × *Laelia* × *Sophranitis* = *Sophranocattleya*; *Cochlioda* × *Miltonia* × *Odontoglossum* = *Vuyllstekeara*.

In mosses, intrageneric hybrids were found or produced artificially for *Funaria*×*Physcomitrium*, *Physcomitrium*×*Physcomitrella*, *Weissia*×*Astomum* (the latter genus is often treated within *Weissia*), *Ditrichum*×*Pleuridium* (see review of Anderson, 1980). Some *Physcomitrium*×*Physcomitrella* hybrids produce tetrads with 2 of 4 spores viable. According to Wettstein (1924) they have only maternal chromosomes, thus not further reproducing hybrid organisms.

Some of the recent phylogenetic studies, with construction of haplotype network by two independent markers illustrate the possible hybridization in the genus *Sciuro-hypnum* (Draper & Hedenäs, 2009).

The case of *Podperaea bairii* likely represents a rare event. ITS is the most variable of DNA regions which are widely used in moss phylogenetics, but not that variable to be useless for phylogeny reconstruction. The large trees that were based on, among others, ITS data, provided many helpful results on the relationships of pleurocarpous mosses (Gardiner et al., 2005; Ignatov et al., 2007; Huttunen et al., 2012).

However the case of *Podperaea bairii* might be important for understanding of a strong incongruence between topology of molecular trees and morphology, like for example, the case of *Pinnatella homaliadelphoides* Enroth, S. Olsson, S. He, Shevock & D. Quandt (Enroth et al., 2009).

## ACKNOWLEDGEMENTS

We are grateful to Profs. Bai X.-L. and Zhao D.-P. for arranging a field trip in China, to E. Ignatova for preparing illustrations of the species. The work was partly supported by RFBR 10-04-91150 and 10-04-00678.

## LITERATURE CITED

- ANDERSON, L.E. 1980. Cytology and reproductive biology of mosses. – In: R.J. Taylor & A.E. Leviton (eds.) *The mosses of North America*. San Francisco. Pacific Div. Am. Assoc. Adv. Sci. 37-76.
- BAI, X.-L. 2010. Bryoflora Helanshanica. *Typis Ningxia Popularis*. Yinchuan. 281 pp. [In Chinese].
- DRAPER, I. & L. HEDENÄS 2009. Circumscription of European taxa within the *Sciuro-hypnum reflexum* complex (Brachytheciaceae, Bryophyta), based on molecular and morphological data. – *Taxon* **58**: 572-584.
- ENROTH J., S. OLSSON, S. HE, J.R. SHEVOCK & D. QUANDT 2010. When morphology and molecules tell us different stories, part 2: Pinnatella homaliadelphoides (Neckeraceae), a new moss species from China and India. – *Tropical Bryology* **31**: 67-75.
- GARDINER, A., M. IGNATOV, S. HUTTUNEN & A. TROITSKY 2005. On resurrection of the families Pseudoleskeaceae Schimp. and Pylaisiaceae Schimp. (Musci, Hypnales). – *Taxon* **54**: 651-663.
- GOLOBOFF, P. A. 1994. *NONA: A Tree Searching Program. Program and documentation*. Argentina, Tucumán, published by the author.
- HALL, T. A. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. – *Nucl. Acids. Symp.* **41**: 95-98.
- HEDENÄS, L. 1997. A partial revision of *Campylium* (Musci). – *Bryologist* **100**: 65-88.
- HUTTUNEN, S., N. BELL, V.K. BOBROVA, V. BUCHBENDER, W.R. BUCK, C.J. COX, B. GOFFINET, L. HEDENÄS, B.-C. HO, M.S. IGNATOV, O. KUZNETSOVA, I.A. MILYUTINA, A. NEWTON, S. OLSSON, L. POKORNY, J.A. SHAW, M. STECH, A. TROITSKY, A. VANDERPOORTEN, D. QUANDT [2012]. Disentangling knots of rapid evolution: origin and diversification of the moss order Hypnales. – *J. Bryol.* (submitted).
- IGNATOV, M.S., H. ANDO & E.A. IGNATOVA 1996. Bryophyte flora of Altai Mountains. VII. Hypnaceae and related pleurocarps with bi- or ecostate leaves. – *Arctoa* **6**: 21-112.
- IGNATOV, M. & L. HEDENÄS 2007. Homologies of stem structures in pleurocarpus mosses, especially of pseudoparaphyllia and similar structures. – In: Newton, A.E. & R. Tangney (eds.), *Pleurocarpus mosses: systematics and evolution*. CRC Press, Boca Raton–London–New York: 269-286.
- IGNATOV, M., A. GARDINER, V. BOBROVA, I. MILYUTINA, S. HUTTUNEN & A. TROITSKY 2007. On relationships of mosses of the order Hypnales, with the special reference to taxa traditionally classified in Leskeaceae. – In: Newton, A.E. & R. Tangney (eds.), *Pleurocarpus mosses: systematics and evolution*. CRC Press, Boca Raton–London–New York: 177-213.
- IWATSUKI, Z. & J.M. GLIME 1984. Podperaea, a new genus of Hypnaceae). – *J. Hattori Bot. Lab.* **55**: 495-500.
- NIXON, K.C. 1999. *Winclada (BETA) ver. 0.9.9*. available at [http://www.cladistics.com/about\\_winc.html](http://www.cladistics.com/about_winc.html).
- OCHYRA, R., J. ŻARNOWIEC & H. BEDNAREK-OCHYRA 2003. Census catalogue of Polish mosses. – *Polish Acad. Sci., Inst. Bot., Krakow*: 372 pp.
- OLSSON, S., V. BUCHBENDER, J. ENROTH, L. HEDENÄS, S. HUTTUNEN, & D. QUANDT 2009. Phylogenetic analyses reveal high levels of polyphyly among pleurocarpus lineages as well as novel clades. – *Bryologist* **112**(3): 447-466.
- PEDERSEN, N. & L. HEDENÄS 2002. Phylogeny of the Plagiotheciaceae Based on Molecular and Morphological Evidence. – *Bryologist* **105**(3): 310-324.
- [TROITSKY, A.V., M.S. IGNATOV, V.K. BOBROVA & I.A. MILYUTINA] ТРОИЦКИЙ, А.В., М.С. ИГНАТОВ, В.К. БОБРОВА, И.А. МИЛЮТИНА 2007. Вклад геносистематики в современное представление о филогении и системе моховидных. – [Contribution of molecular methods to the phylogeny and taxonomy of Bryophyta] *Биохимия [Biochemistry]* **72**(12): 1690-1704.
- VANDERPOORTEN, A. & L. HEDENÄS. 2009. New combinations in the Amblystegieaceae. – *J. Bryol.* **31**: 129-132.
- WETTSTEIN, F. v. 1924. Kreuzungsversuche mit multiploiden Moosrasen. – *Biol. Zentralbl.* **44**: 145-168.
- ZHAO, D.-P., X.-L. BAI, X.-D. WANG & H.-M. JING 2006. Bryophyte Flora of Helan Mountain in China. – *Arctoa* **15**: 219-235.

## Appendix 1. Studied specimen data and GenBank accessions numbers.

Species	Specimen	GenBank assession
<i>Entodon seductrix</i>	U.S.A., New Jersey, Tan #92-129 (MHA)	JN896314
<i>Herzogiella turfacea</i>	Russia, Kunashir, Ignatov #06-1262 (MW)	JN896315
<i>Homomallium incurvatum</i>	Russia, Altai, coll. Ignatov 0/1714 (MHA)	JQ247725
<i>Homomallium connexum</i>	Russia, Primorsky Territory, Ignatov #07-572 (MHA)	JN896316
<i>Hookeria lucens</i> (Hedw.) Sm.	U.S.A., California, Ignatov 89-1 (MHA)	JN896317
<i>Isopterygiopsis muelleriana</i> (Schimp.) Z. Iwats. 2	Russia, Primorsky Territory, Ignatov #07-296 (MHA)	JN896318
<i>Mittenothamnium reptans</i>	Honduras, Allen # 12298 (MHA)	JN896320
<i>Ochyraea tatrensis</i> Váňa	Czechoslovakia, coll. Váňa s. n. 30.VIII.1987 (MHA)	JQ247724

<i>Podperaea baii</i> Xilin Gol 1 (gametophyte)	China, Inner Mongolia, Xilin Gol, Bai X.-L. 058141 (HIMC, MHA)	JN896321
<i>Podperaea baii</i> Xilin Gol 2 (clone 1 from gametophyte)	China, Inner Mongolia, Xilin Gol, Bai X.-L. 058141 (HIMC, MHA)	JN896322
<i>Podperaea baii</i> Xilin Gol 3 (sporophyte)	China, Inner Mongolia, Xilin Gol, Bai X.-L. 058141 (HIMC, MHA)	JN896323
<i>Podperaea baii</i> Hohhot	China, Inner Mongolia, Hohhot Kou Jin #110(7) (HIMC, MHA)	JN896324
<i>Podperaea baii</i> Manhan	China, Inner Mongolia, Manhan Shan Ignatov #11-1 (MHA)	JN896325
<i>Podperaea krylovii</i> Altai	Russia, Altai, Ignatov #1/10 (MHA)	JN896326
<i>Podperaea krylovii</i> Khingan	China, Inner Mongolia, Great Khingan Mt., Bai Xue-liang 04095 (MHA)	JN896327
<i>Podperaea krylovii</i> Primorsky 1	Russian Far East, Primorsky Territory Ignatov #08-308 (MHA)	JN896328
<i>Podperaea krylovii</i> Primorsky 2	Russia, Primorsky Territory, Ignatov #07-220 (MHA)	JN896329
<i>Stereodon fauriei</i>	Russia, Khabarovsk Territory, Ignatov, #97-216 (MHA)	JN896313

### B. Previous sequences used in the present analysis

ITS from previous studies (specimen data in Ignatov et al., 2007 or else); if ITS1 and ITS2 have different accessions they are given with /; the only accessions means both ITS1&2, unless otherwise is specially marked.

*Breidleria pratense* (J. Koch ex Spruce) Loeske AY693659; *Calliergonella cuspidata* (Hedw.) Loeske AF168145; *Campylidium sommerfeltii* (Myr.) Ochyra AY693654; *Campylium stellatum* (Hedw.) C. Jens. AF168151; *Campylophyllum halleri* (Sw. ex Hedw.) M. Fleisch. (1) AF168134; *Campylophyllum halleri* (Sw. ex Hedw.) M. Fleisch.(2) AY693655; *Caribaeohypnum polypterum* (Mitt.) Ando & Higuchi AY009799; *Ctenidium molluscum* (Hedw.) Mitt. AF230989 / AF403632; *Drepanium (Hypnum) recurvatum* (Lindb. & Arnell) G. Roth AY693660; *Entodontopsis leucostega* (Brid.) W. R. Buck & R. R. Ireland AY999175; *Eurohypnum leptothallum* (Müll. Hal.) Ando AY695733 / AY695786; *Fabronia ciliaris* (Brid.) Brid. AY528883; *Glossadelphus ogatae* Broth. & Yas. AY999169; *Herzogiella seligeri* (Brid.) Z. Iwats. AY999174; *Heterocladium dimorphum* (Brid.) Bruch & al. AY695757 / AY695771; *Hygrohypnella ochracea* (Wils.) Ignatov & Ignatova AF168138; *Hygrohypnella polaris* (Lindb.) Ignatov & Ignatova AY695735; *Hylocomium splendens* (Hedw.) Bruch et al. AJ288336 / AJ270021; *Hypnum cupressiforme* Hedw. AY528888 / AF403607; *Isopterygiopsis muelleriana* (Schimp.) Z. Iwats. 1 AY528882; *Isopterygiopsis pulchella* (Hedw.) Z. Iwats. AY695751 / AY695770; *Leptodictyum riparium* (Hedw.) Warnst. AF168163; *Leptopterigynandrum austro-alpinum* Müll. Hal. (1) AY693656; *Leptopterigynandrum austro-alpinum* Müll. Hal. (2) AF516163 / AF516158; *Lescuraea secunda* Arnell AF516164 / AF516150; *Myrinia pulvinata* (Wahlenb.) Schimp. AY528886 / AY528887; *Myrinia rotundifolia* (Arnell) Broth. AY528898; *Myurella sibirica* AJ288415 / AJ277227; *Neckera pennata* Hedw. AY009809; *Ochyraea norvegica* (Bruch & al.) Ignatov & Ignatova AY695753 / AY695789; *Ochyraea mollis* (Hedw.) Ignatov AY999178; *Plagiothecium denticulatum* (Hedw.) Bruch & al. AY528892 / AY528893; *Platydictya jungermannioides* (Brid.) H. A. Crum AF168162; *Pylaisia polyantha* (Hedw.) Schimp. AY528881; *Rhytidiopsis robusta* (Hook.) Broth. AJ288331 / AJ288545; *Sanionia uncinata* (Hedw.) Loeske AF168148; *Scorpidium scorpioides* (Hedw.) Limpr. AY009790; *Scorpidium cossonii* AY625996; *Struckia argentata* Yunnan DQ836734; *Struckia argentata* Nepal Q836733; *Struckia enervis* Sichuan DQ836736 / DQ836737; *Struckia enervis* Altai DQ836735; *Stereodon (Hypnum) vaucheri* (Lesq.) Lindb. ex Broth. AY695762 / AY695791; *Stereophyllum radiculosum* (Hook.) Mitt. AY999176; *Taxiphyllum wissgrillii* (Garov.) Wijk & Marg. AY999168; *Tomentypnum falcifolium* (Renauld ex Nichols) Tuom. AF168136; *Tomentypnum nitens* (Hedw.) Loeske AF168161.