

SIMPLE SEQUENCE REPEATS MINING USING COMPUTATIONAL APPROACH IN  
CHLOROPLAST GENOME OF *MARCHANTIA POLYMORPHA*

ПОИСК ПРОСТЫХ НУКЛЕОТИДНЫХ ПОВТОРОВ В ХЛОРОПЛАСТНОМ ГЕНОМЕ  
*MARCHANTIA POLYMORPHA* С ПОМОЩЬЮ КОМПЬЮТЕРНОЙ ПРОГРАММЫ

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Abstract

Simple sequence repeats (SSRs) or microsatellites are found in DNA sequences and consist of short repeat motifs of 1-6 bp. These repeats play important role in the development of molecular markers, phylogenetics, population genetics and evolutionary biology. The present analysis was conducted to detect chloroplastic SSRs (cpSSRs) in *Marchantia polymorpha*. The chloroplast genome sequence of *M. polymorpha* was downloaded from the National Center for Biotechnology Information (NCBI) and mined with the help of a Perl script named MISA. A total of 66 cpSSRs were detected in 121.024 kb sequence mined showing density of 1 SSR/1.83 kb. Depending on the repeat units, the length of SSRs ranged from 12 to 17 bp for mono, 12 to 64 bp for di, 12 to 21 bp for tri, 12 to 24 bp for tetra, 15 bp for penta and 18 bp for hexa nucleotide repeats. Mononucleotide repeats were the most frequent repeat type (42.42%) followed by dinucleotide (25.76%) and tetranucleotide (21.21%) repeats. PCR primers were successfully designed for 45 (68.18%) cpSSRs of *M. polymorpha*.

Резюме

Простые нуклеотидные повторы (SSRs), или микросателлиты, встречающиеся в последовательностях ДНК, состоят из коротких повторов из 1-6 нуклеотидов. Они важны для разработки молекулярных маркеров для филогенетических, популяционно-генетических и эволюционных исследований. Представленный анализ проведен с целью определения хлоропластных SSR *Marchantia polymorpha*. Использован хлоропластный геном *M. polymorpha* из ГенБанка и в нем проведен поиск с помощью оригинального скрипта MISA на языке Perl. Выявлено 66 SSR маркеров в последовательности длиной 121024 bp; таким образом, имеется 1 SSR повтор на 1.83 kbp. Длина SSR повторов варьирует от 12 до 17 bp для моно-, 12 до 64 bp для ди-, 12 до 21 bp для три-, 12 до 24 bp для тетра-, 15 bp для пента- и 18 bp для гекса-нуклеотидных повторов. Мононуклеотидные повторы наиболее часты (42.42%), реже встречаются ди- (25.76%) и тетра- нуклеотидные повторы (21.21%). Праймеры успешно разработаны для 45 (68.18%) SSR маркеров.

KEYWORDS: Bryophytes, Marchantia, Chloroplast, Microsatellites, Simple Sequence Repeats

INTRODUCTION

Bryophytes are broadly classified into liverworts, mosses and hornworts, and are considered as the earliest land plants. Chloroplast and mitochondrial genome sequences based phylogenetic analysis showed liverworts as the earliest diverging lineage and hornworts as sister group to vascular plants (Shanker, 2013a,b,c).

Among land plants the first complete chloroplast genome sequence determined was that of *Marchantia polymorpha* (Ohyama *et al.*, 1986; Ohyama *et al.*, 1988). Since then several studies to elucidate the structure and organization of chloroplast genomes have been conducted. However, a small number of organelle genome sequences of bryophytes are available (Shanker, 2012a,b).

Simple sequence repeats (SSRs) or microsatellites are found in DNA sequences and consist of short repeat motifs of 1-6 bp (Shanker *et al.*, 2007b). These

repeats are present in both coding and non-coding regions of DNA sequences, and have been widely used as molecular markers in plant genomes (Gupta *et al.*, 2003; Jakobsson *et al.*, 2007; Blair & Hurtado, 2013). Studies were conducted to mine SSRs in chloroplast genomes of bryophytes (Shanker, 2013d,e). However, mining of other available organelle genome sequences of bryophytes will help to know the occurrence of SSRs in them.

Biotechnological methods of SSR identification are costly and consume time. Contrary to this bioinformatic approaches offer rapid and economical SSR extraction using sequences deposited in public databases (Shanker *et al.*, 2007a). Therefore the present analysis was conducted to identify cpSSRs in *M. polymorpha*. Moreover, the distribution of these repeats in coding and non-coding regions of chloroplast genome was analyzed.

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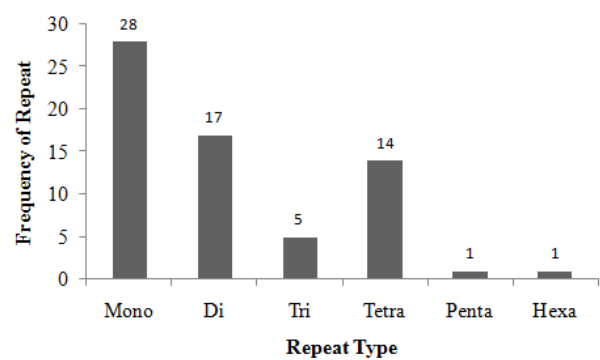


Fig. 1. Frequency distribution of mono-hexa repeats.

#### MATERIALS AND METHODS

##### *Chloroplast genome sequence of M. polymorpha.*

The complete chloroplast genome sequence of *M. polymorpha* (NC\_001319, 121024 bp; Ohyama *et al.*, 1986) was downloaded from NCBI ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)) in FASTA and GenBank format.

**Identification of chloroplastic simple sequence repeats.** A Perl script named MISA (available at <http://pgrc.ipkgatersleben.de/misa/misa>) was used to identify SSRs in chloroplast genome sequence of *M. polymorpha*. MISA takes FASTA formatted DNA sequence file as an input and generates information of perfect and compound SSRs, if found. A perfect SSR consist of same repeating motif without interruptions, e.g., (GTG)<sub>8</sub>. In compound SSR, two or more SSRs are found adjacent to one another, e.g., (GTG)<sub>8</sub>(AT)<sub>16</sub> (Bachmann & Bare, 2004). The length of SSRs in this study was defined as  $\geq 12$  for mono, di, tri and tetranucleotide,  $\geq 15$  for pentanucleotide and  $\geq 18$  for hexanucleotide repeats. Maximum difference between two compound SSRs was taken as 0. Since the information of coding and non-coding regions of chloroplast genome has been available in GenBank file, therefore, the mined cpSSRs were classified as coding, non-coding and coding-non-coding (occurrence of few bases of SSR in coding as well as in non-coding regions or vice-versa) SSRs based on the presence of these repeats in respective regions.

**Primer designing for identified SSRs.** PCR primers were designed for identified SSRs considering 200 base pair of SSR flanking regions using Primer 3 (<http://bioinfo.ut.ee/primer3-0.4.0/>) with default parameters of GC content, melting temperature, primer and PCR product size.

#### RESULTS AND DISCUSSION

In this study SSRs were identified in chloroplast genome sequence of *M. polymorpha* considering a minimum length of 12 bp. A total of 66 perfect SSRs were detected with length variation from 12 to 64 bp. Compound SSRs were found to be totally absent in chloroplast genome sequence of *M. polymorpha*. The frequency of identified SSRs (mono-hexa) is presented in Fig. 1. Detail information of mined SSRs motif, their length, start-end position and the region in which they lie is presented in Table 1. It is evident from this table that out of total cpSSRs detected, 14 (21.21%) found in coding, 48 (72.73%) in non-coding and only 4 (6.06%) in coding-

non-coding regions. The occurrence of these repeats in various regions attributed to the evolutionary processes that fine tune distribution of SSR repeat types in genome (Lin & Kussell, 2012).

Mononucleotides were found to be most frequent repeat (28, 42.42%) followed by dinucleotide (17, 25.76%), tetranucleotide (14, 21.21%) and trinucleotide (5, 7.58%) repeats. Penta and hexanucleotide repeats were found with equal frequencies (1, 1.52%). An attempt was made to design PCR primers for all mined SSRs, however, these were successfully designed for 45 (68.18%) cpSSRs of *M. polymorpha*. A list of designed PCR primers, their length, product size etc. is presented in Table 2.

The mined SSRs represent a density of 1 SSR/1.83 kb in 121.024 kb sequence mined. The density of cpSSRs in *M. polymorpha* found to be higher than the density of cpSSRs in *Anthoceros formosae* (1 SSR/2.4 kb; Shanker, 2013d), rice (1 SSR/6.5 kb; Rajendrakumar *et al.*, 2007), EST-SSRs in barley, maize, wheat, rye, sorghum and rice (1 SSR/6.0 kb; Varshney *et al.*, 2002), cotton and poplar (1 SSR/20 kb and 1 SSR/14 kb respectively; Cardle *et al.*, 2000), Unigenes sequences of *Citrus* (1 SSR/12.9 kb; Shanker *et al.*, 2007a). However, the density of cpSSRs in *M. polymorpha* found to be lower than the cpSSRs density in family Solanaceae (1 SSR/1.26kb; Tambarussi *et al.*, 2009). The selection of SSR detection tools, parameters taken (e.g. minimum length of SSRs) and amount of data analyzed might be the cause of variations in SSR density.

The higher occurrence of mononucleotides in this study shows consistency with earlier studies of cpSSRs in *Anthoceros formosae* (Shanker, 2013d), rice (Rajendrakumar *et al.*, 2007), Solanaceae species (Tambarussi *et al.*, 2009), *Saccharum* spp. (Melotto-Passarini *et al.*, 2011) and *Olea* species (Filiz & Koc, 2012). Moreover the abundance of cpSSRs in non-coding regions is also in agreement with previous studies (Hancock, 1995; Shanker, 2013d).

As a concluding remark, *in silico* mining of complete chloroplast genome sequence of *M. polymorpha* saves time, cost and provides sufficient number of SSRs for this liverwort. The identified SSRs can be used to develop SSR markers and for various other purposes.

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No.	MOTIF	LENGTH	START	END	REGION
1	(ATAAA)3	15	450	464	Non coding
2	(AT)7	14	2290	2303	Non coding
3	(TAA)4	12	2510	2521	Non coding
4	(TA)32	64	3589	3652	Non coding
5	(AAC)7	21	4559	4579	Non coding
6	(AT)6	12	5318	5329	Non coding
7	(TA)17	34	5379	5412	Non coding
8	(T)12	12	5491	5502	Non coding
9	(A)14	14	7640	7653	Coding
10	(T)14	14	9981	9994	Non coding
11	(A)15	15	13493	13507	Coding
12	(A)12	12	14664	14675	Coding
13	(A)13	13	15824	15836	Coding
14	(T)12	12	22002	22013	Non coding
15	(AT)24	48	22693	22740	Non coding
16	(AAT)6	18	23325	23342	Non coding
17	(A)13	13	23923	23935	Non coding
18	(T)13	13	26530	26542	Non coding
19	(AAAC)3	12	26632	26643	Non coding
20	(AT)7	14	26664	26677	Non coding
21	(T)12	12	26745	26756	Non coding
22	(A)15	15	30115	30129	Coding
23	(A)13	13	33266	33278	Coding
24	(ATTA)3	12	36685	36696	Non coding
25	(TA)8	16	38168	38183	Non coding
26	(A)13	13	38339	38351	Non coding
27	(TTAA)6	24	41278	41301	Coding-Non-Coding
28	(A)12	12	41323	41334	Non coding
29	(TA)13	26	41988	42013	Non coding
30	(A)16	16	50459	50474	Non coding
31	(TAA)4	12	50760	50771	Non coding
32	(AT)6	12	53692	53703	Non coding
33	(ATTA)3	12	53937	53948	Non coding
34	(TA)8	16	56040	56055	Non coding
35	(AT)16	32	56076	56107	Non coding
36	(A)12	12	58041	58052	Non coding
37	(TTA)4	12	59082	59093	Non coding
38	(A)13	13	60479	60491	Coding
39	(TTTG)3	12	63125	63136	Coding
40	(T)12	12	63943	63954	Non coding
41	(TA)6	12	64048	64059	Non coding
42	(A)16	16	64355	64370	Coding-Non-Coding
43	(TTATAT)3	18	64657	64674	Non coding
44	(AT)6	12	64681	64692	Non coding
45	(T)13	13	64817	64829	Non coding
46	(TA)6	12	64853	64864	Non coding
47	(TA)7	14	67473	67486	Non coding
48	(TATT)4	16	68702	68717	Non coding
49	(TAAA)3	12	70571	70582	Non coding
50	(T)13	13	72980	72992	Non coding
51	(T)12	12	75959	75970	Coding
52	(TTTA)3	12	76223	76234	Non coding
53	(T)15	15	76621	76635	Coding-Non-Coding
54	(T)13	13	77537	77549	Non coding
55	(TTTA)3	12	79756	79767	Non coding
56	(T)13	13	79811	79823	Non coding
57	(T)17	17	80642	80658	Coding
58	(AGGT)3	12	87572	87583	Coding
59	(TTTA)3	12	95400	95411	Non coding
60	(AATA)3	12	96864	96875	Coding
61	(AT)6	12	98203	98214	Non coding
62	(AT)7	14	98579	98592	Non coding
63	(A)16	16	99738	99753	Non coding
64	(AATT)3	12	99773	99784	Coding-Non-Coding
65	(T)14	14	106439	106452	Coding
66	(CTAC)3	12	114535	114546	Coding

Table 1: Information of mined SSRs in chloroplast genome sequence of *M. polymorpha*.

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Table 2: List of primers designed for identified SSRs along with primer length, melting temperature and product size.

No.	MOTIF	START	END	LEFT / RIGHT PRIMER	PRIMER LENGTH	T <sub>m</sub>	GC%	PRODUCT SIZE (bp)
1	(AT) <sub>7</sub>	2290	2303	ATGAAGGAGTGCGATTCGTT AAAAATTAATGGTGAAATTTGCGTT	20 27	57.613 57.205	45 22.222	152
2	(TAA) <sub>4</sub>	2510	2521	AACGCAAAATTCACCAATTAATTTT TCGAAGAATCCTTCTATTCAATGGT	27 26	57.205 58.661	22.222 34.615	261
3	(TA) <sub>17</sub>	5379	5412	GCCCAGGCTATATTAATAATATCCATT AGACAACCCATCATAAGATTAACA	27 26	57.186 57.122	33.333 30.769	300
4	(A) <sub>14</sub>	7640	7653	ACTGTCTCATCGCATGGAGG AGCAGCTCCGCTGCTAAAA	20 20	59.537 59.678	55 50	196
5	(T) <sub>14</sub>	9981	9994	TCGAAATAGAGCAAAAACCTAAAGGA CGTCATCCGGCTCAAATAGT	26 20	59.062 57.775	34.615 50	250
6	(AT) <sub>24</sub>	22693	22740	TGACCTCAATTTTCTAGATAGTCAAATT CACTCGGCCATCTCTCCAAA	26 20	57.131 59.75	30.769 55	393
7	(AAT) <sub>6</sub>	23325	23342	TGCAATTGATTACTACTTTGCCA GCCTTTGTATGGCAAGCTTCA	23 21	57.026 59.454	34.783 47.619	220
8	(A) <sub>13</sub>	23923	23935	CGAATCCTTCCGTCCAGAC GCAGGACCAGCATACATCCA	20 20	60.179 59.819	60 55	231
9	(T) <sub>13</sub>	26530	26542	TGTGTCTAGAAATAACCAGCATGC TGTTTGTTTGTTTTTCAAAGTAGGCA	24 27	59.364 59.773	41.667 29.63	192
10	(AAAC) <sub>3</sub>	26632	26643	AGAAATAACCAGCATGCTGATTAAT TGCAATCCATAAAAAATTTCTGAA	25 26	57.407 57.085	32 26.923	250
11	(AT) <sub>7</sub>	26664	26677	AAAGTTTTGCCTACTTTTGAAAAACA TGCAATCCATAAAAAATTTCTGAA	26 26	57.615 57.085	26.923 26.923	100
12	(ATTA) <sub>3</sub>	36685	36696	CCAGCGATTTACAGTCCGT TCTCTTCAAGGAGGCGACG	20 20	60.108 59.756	55 55	163
13	(TA) <sub>8</sub>	38168	38183	TCCAATAGGTTATCAATCTTTCCGA CACTGAGTTAAAAGGGCAATAAGA	25 24	57.868 57.182	36 37.5	387
14	(TTAA) <sub>6</sub>	41278	41301	CTGCTGCAGCTGGTTTTGAA TCGAGGGTTCAAATCCCTCTC	20 21	59.614 59.168	50 52.381	223
15	(A) <sub>12</sub>	41323	41334	CTGCTGCAGCTGGTTTTGAA TCGAGGGTTCAAATCCCTCTC	20 21	59.614 59.168	50 52.381	223
16	(A) <sub>16</sub>	50459	50474	GCATTACAAGTGCACGCTC CCAATTCGCCATACCCCT	20 20	60.247 60.106	55 55	249
17	(TAA) <sub>4</sub>	50760	50771	ACAAAAGAGGTGCAGAGACTCA CTGGGGGTAGAGGGACTTGA	22 20	59.564 59.957	45.455 60	248
18	(AT) <sub>6</sub>	53692	53703	TACCGCTGAGCTATAGCCCT CTCTCGCGTATGAAAGCGA	20 20	59.887 60.248	55 55	216
19	(ATTA) <sub>3</sub>	53937	53948	TCGCTTTCATACGGCGAGAG AGAGCTAAAGCAAGATTAGAAGCA	20 24	60.248 58.261	55 37.5	176
20	(A) <sub>12</sub>	58041	58052	TGGGTTTGTAGCTCAGTGGA AGCGCCAATTAATCCACCAA	20 20	58.57 58.154	50 45	247
21	(TTA) <sub>4</sub>	59082	59093	TGAATTATATAACGCTGCTCCTTGT GCAGGAAAAATTAATCCAATAAGGA	25 27	58.712 58.689	36 33.333	292
22	(A) <sub>13</sub>	60479	60491	TCAAAACGTTCTTGCGAGTCT AGCTCTAATCCAAGCTAATTTTCTGT	21 26	58.361 58.885	42.857 34.615	235
23	(TTTG) <sub>3</sub>	63125	63136	ACCCCAATAGAGGCTAGTACGA ACCGTAAGATGGTTAGCCGT	22 20	59.824 58.811	50 50	175
24	(TA) <sub>6</sub>	64048	64059	TTACAGATTGGTCTATCCGAAA ACGCTAATGTAAAGCACCGA	23 21	57.057 58.309	39.13 42.857	345
25	(A) <sub>16</sub>	64355	64370	TCGGTGCTTTAACATTAGCGT TCCAGACAACAAAGCTTCAACC	21 22	58.309 59.312	42.857 45.455	209
26	(TTATAT) <sub>3</sub>	64657	64674	TGGAGACCTACGTTCTACCGA TGGTAGCGCGTTTGTTTTGG	21 20	59.721 59.97	52.381 50	183
27	(AT) <sub>6</sub>	64681	64692	TGGAGACCTACGTTCTACCGA TGGTAGCGCGTTTGTTTTGG	21 20	59.721 59.97	52.381 50	183
28	(TA) <sub>6</sub>	64853	64864	CCAACTGCGTACATCCCT ACAGGTGCAGTAGAAAGATATGT	20 23	60.393 57.316	55 39.13	297
29	(TATT) <sub>4</sub>	68702	68717	ACTTTCGGAACCAATAGGCA ACAATGGAGAGATTGGTCCCA	22 21	60.225 58.722	45.455 47.619	233



Table 2 (cont.): List of primers designed for identified SSRs along with primer length, melting temperature and product size.

No.	MOTIF	START	END	LEFT / RIGHT PRIMER	PRIMER LENGTH	T <sub>m</sub>	GC%	PRODUCT SIZE (bp)
30	(TAAA)3	70571	70582	GCTCAAGTGGGAATTTGGAGCG TGATTCCTAAAGTACCTACCAACA	21 24	60.136 57.061	52.381 37.5	232
31	(T)13	72980	72992	TGAATAGTAAAGACTTCGTAAATCCA TCATCCGGCTCCAACATTAATGA	27 23	57.068 60.118	29.63 43.478	280
32	(T)12	75959	75970	CCCATTCCACCTAAAACCTTTTGGGA AACAGTTC AAGTACCTGCTACT	24 22	59.409 57.042	41.667 40.909	241
33	(TTTA)3	76223	76234	TCGCAATTGTATCATTCCCCA AGGAACTCGAGTTTTTGGTCCA	21 22	57.711 59.828	42.857 45.455	192
34	(T)15	76621	76635	TGGCACTGCTTCTTTAACAACA GGCCCTCCCTAATCCATCCA	22 20	58.98 60.772	40.909 60	195
35	(T)13	77537	77549	TGCGTTTTAATACTATAACAAAAACCC GCTATGCTTAGTGTGTGACTCG	27 22	57.377 59.145	29.63 50	199
36	(T)13	79811	79823	TCATTTAGCATCAGAAAGGATGACT AGGAAAAATAGGCAAGTAAAAAGGT	25 26	58.343 59.9	36 34.615	277
37	(T)17	80642	80658	ACAGTATATCCTGTTGTCGTACCT TTATGAATCAAGTTAAGTACCCAGTAC	24 27	58.801 57.12	41.667 33.333	212
38	(AGGT)3	87572	87583	CTTCCAGCCAATGTCCGAGT TTATCTCGCGCCCTAGGTA	20 20	60.036 59.889	55 55	153
39	(TTTA)3	95400	95411	AGCAGTGCTAAGGCTTCTCG ACGCTCTAAGGTTATAAATGGCA	20 23	60.108 57.969	55 39.13	194
40	(AATA)3	96864	96875	TTTCTCGTGGTCCAGCATCC GCTTTGCCAGGTATGAGTGG	20 20	60.036 58.902	55 55	227
41	(AT)6	98203	98214	ACGGGATTACTAAACCTGCAGA TGAGACTACTCGTAGTATGGGTCT	22 24	59.164 59.592	45.455 45.833	226
42	(A)16	99738	99753	TCTGGTAATTCATAAAGCTCACAA CCTAAAGGCAAAATAGAAGGGCA	25 23	57.006 58.977	32 43.478	165
43	(AATT)3	99773	99784	TCTGGTAATTCATAAAGCTCACAA CCTAAAGGCAAAATAGAAGGGCA	25 23	57.006 58.977	32 43.478	165
44	(T)14	106439	106452	TGGAAAATCCCATCTGTTTGCT AGGATCTATGCGTGCTCGAC	22 20	58.491 59.688	40.909 55	248
45	(CTAC)3	114535	114546	TTATCTCGCGCCCTAGGTA CTTCCAGCCAATGTCCGAGT	20 20	59.889 60.036	55 55	153